

线粒体动力学失衡与新型冠状病毒感染相关急性呼吸窘迫综合征的研究进展

张子佳¹ 杜斌¹ 吴逊尧¹ 胡晓芸² 刁世童¹ 董润¹

¹ 北京协和医院 MICU, 北京 100010; ² 北京协和医院重症医学科, 北京 100010

通信作者: 杜斌, Email: dubin98@gmail.com

【摘要】 新型冠状病毒(新冠病毒)感染所致重症肺炎患者常合并急性呼吸窘迫综合征(ARDS),病死率高。ARDS 以弥漫性肺泡损伤、肺水肿和低氧血症为特征,线粒体在低氧和病毒感染下易出现形态及功能异常,进而引发细胞凋亡和损伤,严重影响疾病进程。线粒体通过裂变和融合维持稳态,ARDS 时低氧使动力相关蛋白 1(Drp1)磷酸化,引发线粒体过度分裂,破坏肺泡上皮屏障。动物实验显示抑制该过程可减轻肺损伤,为治疗提供潜在方向。新冠病毒感染相关 ARDS 的病理机制与典型 ARDS 类似,但程度更严重,病毒感染和低氧导致线粒体平衡被打破,引发裂变和自噬异常,促使氧化应激和线粒体 DNA(mtDNA)释放,激活炎症小体,诱导缺氧诱导因子-1 α (HIF-1 α)表达,加剧病毒感染、炎症和凝血反应,导致多器官损伤。针对新冠病毒感染相关 ARDS 的治疗常使用机械通气和糖皮质激素。机械通气易导致肺和膈肌损伤、线粒体动力学改变,肺保护性通气策略可减少不良影响;糖皮质激素可调节线粒体功能和免疫反应,通过多途径改善病情。新冠病毒感染相关线粒体动力学失衡源于低氧和病毒蛋白,引发肺部及多器官损伤。本综述旨在明确线粒体动力学失衡与新冠病毒感染相关 ARDS 的病理生理学机制,探索调控线粒体动力学平衡以治疗该疾病的有效策略,为新冠病毒感染相关 ARDS 患者提供新的治疗靶点与方法。由于现有治疗有局限,未来需深入研究线粒体功能障碍机制,开发新疗法和调控策略,以提升治疗效果。

【关键词】 线粒体动力学; 新型冠状病毒感染; 急性呼吸窘迫综合征

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Research progress on the relationship between mitochondrial dynamics imbalance and novel coronavirus infection-related acute respiratory distress syndrome

Zhang Zijia¹, Du Bin¹, Wu Xunyao¹, Hu Xiaoyun², Diao Shitong¹, Dong Run¹

¹MICU, Peking Union Medical College Hospital, Chinese Academy of Medical Sciences & Peking Union Medical College, Beijing, 100010, China; ²Department of Critical Care Medicine, Peking Union Medical College Hospital, Beijing, 100010, China
Corresponding author: Du Bin, Email: dubin98@gmail.com

【Abstract】 Patients with severe pneumonia caused by novel coronavirus infection are often complicated with acute respiratory distress syndrome (ARDS), which has a high mortality. ARDS is characterized by diffuse alveolar damage, pulmonary edema, and hypoxemia. Mitochondria are prone to morphological and functional abnormalities under hypoxia and viral infection, which can lead to cell apoptosis and damage, severely impacting the disease progression. Mitochondria maintain homeostasis through fission and fusion. In ARDS, hypoxia leads to the phosphorylation of dynamin-related protein 1 (Drp1), triggering excessive mitochondrial fission and damaging the alveolar epithelial barrier. Animal experiments have shown that inhibiting this process can alleviate lung injury, providing a potential direction for treatment. The pathology of novel coronavirus infection-related ARDS is similar to that of typical ARDS but more severe. Viral infection and hypoxia disrupt the mitochondrial balance, causing fission and autophagy abnormalities, promoting oxidative stress and mitochondrial DNA (mtDNA) release, activating inflammasomes, inducing the expression of hypoxia-inducible factor-1 α (HIF-1 α), exacerbating viral infection, inflammation, and coagulation reactions, and resulting in multiple organ damage. Mechanical ventilation and glucocorticoids are commonly used in the treatment of novel coronavirus infection-related ARDS. Mechanical ventilation is likely to cause lung and diaphragm injuries and changes in mitochondrial dynamics, while the lung protective ventilation strategy can reduce the adverse effects. Glucocorticoids can regulate mitochondrial function and immune response and improve the patient's condition through multiple pathways. The mitochondrial dynamics imbalance in novel coronavirus infection-related ARDS is caused by hypoxia and viral proteins, leading to lung and multiple organ injuries. To clarify the pathophysiological mechanism of mitochondrial dynamics imbalance in novel coronavirus infection-related ARDS and explore effective strategies for regulating mitochondrial dynamics balance to treat this disease, so as to provide new treatment targets and methods for patients with novel coronavirus infection-related ARDS. The existing treatments have limitations. Future research needs to deeply study the mechanism of mitochondrial dysfunction, develop new therapies and regulatory strategies, and improve the treatment effect.

【Key words】 Mitochondrial dynamics; Novel coronavirus infection; Acute respiratory distress syndrome

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急性呼吸窘迫综合征 (acute respiratory distress syndrome, ARDS) 是重症新型冠状病毒 (新冠病毒) 感染患者最常见的并发症, 发生率为 30% ~ 60%, 病死率达 50% ~ 70%^[1], 其特征为弥漫性肺泡损伤、持续炎症反应刺激、肺泡-毛细血管屏障损伤和肺水肿, 临床表现为严重的低氧血症和双侧肺浸润, 最终导致呼吸衰竭^[2-3]。线粒体是动态的细胞器, 不断经历融合/裂变以维持正常的细胞生理活动和能量代谢。低氧和病毒感染均可以诱发线粒体动力学的失衡, 继而诱发线粒体自噬异常、氧化应激和能量代谢紊乱, 最终导致细胞的凋亡和损伤。因此, 本文将以线粒体动力学失衡为切入点, 系统阐述新冠病毒感染相关 ARDS 继发的线粒体动力学失衡所导致的肺部疾病及肺损伤的机制, 并探索重症新冠病毒感染相关 ARDS 的新治疗靶点及策略。

1 线粒体动力学与 ARDS

线粒体是细胞进行有氧呼吸和能量代谢的重要场所, 被称为“能量工厂”, 也是缺血缺氧后即刻发生损伤的细胞器之一。线粒体在细胞内并非静止不动, 而是通过不断地裂变和融合, 维持自身功能的完整性, 并适应细胞内的能量需求, 即线粒体动力学^[4]。线粒体的分裂、融合和转运过程失去平衡时, 可导致线粒体形态、数量、分布及功能异常, 发生线粒体动力学失衡^[5]。线粒体的动力学平衡对于维持线粒体的质量和功能起重要作用。线粒体融合可增加线粒体嵴的重塑并维持线粒体网络的稳定, 融合过程主要由视神经萎缩蛋白 1 (optic atrophy 1, OPA1) 和线粒体融合蛋白 (mitochondrial fusion proteins, Mfn1、Mfn2) 参与^[6]; 线粒体分裂可调节细胞代谢和能量供应, 主要由动力相关蛋白 1 (dynamitin-related protein 1, Drp1) 调控^[7]。

ARDS 是一种急性弥漫性炎症性肺损伤, 由于严重感染、休克、创伤等因素导致肺泡毛细血管膜损伤和通透性增加, 引起肺水肿和肺不张, 从而导致通气肺组织减少。肺泡-毛细血管屏障功能损伤是 ARDS 的重要病理生理学特征, 其临床表现为顽固性低氧血症和进行性呼吸窘迫。线粒体作为细胞的主要耗氧细胞器, 主要通过氧化磷酸化产生三磷酸腺苷 (adenosine triphosphate, ATP) 供能。当细胞内处于持续低氧条件下时, ATP 的生成减少, 细胞会启动代偿机制, 如增强糖酵解途径, 产生较少的 ATP, 并伴随乳酸等副产物的增加。低氧会通过抑制氧化磷酸化调控线粒体动力学平衡、破坏线粒体的完整性, 并诱导 II 型肺泡上皮细胞发生凋亡^[8-9]。此外, 肺内外致病因素可通过引发炎症反应促使肺泡上皮细胞形成不利的内环境, 导致线粒体自噬功能紊乱, 而过度的线粒体自噬激活可加重炎症反应^[10]。低氧环境会使 Drp1 磷酸化水平增加, 促进线粒体发生过度分裂, 磷酸化的 Drp1 同时被招募到线粒体内膜上形成多聚螺旋结构, 收缩挤压线粒体呈现片段化, 使细胞功能受损甚至发生死

亡^[11]。脂多糖 (lipopolysaccharide, LPS) 通过诱导 Drp1 蛋白在 Ser616 位点发生磷酸化, 激活 Drp1 介导的线粒体超分裂, 促进肺泡上皮间紧密连接结构破坏和上皮细胞凋亡, 破坏肺上皮屏障的完整性^[12]。基于 ARDS 小鼠模型的研究显示, 抑制 Drp1 介导的线粒体分裂可以显著减轻 ARDS 肺泡上皮细胞损伤和功能破坏, 提示 ARDS 患者的肺泡上皮细胞发生线粒体动力学失衡, 或可作为临床治疗的潜在靶点^[13]。

2 线粒体动力学失衡与新冠病毒感染相关 ARDS

2.1 新冠病毒感染相关 ARDS: 弥漫性肺泡损伤及急性纤维索性机化性肺炎是新冠病毒感染相关 ARDS 的病理基础。新冠病毒感染相关 ARDS 与典型 ARDS 相似, 组织病理学特征均为弥漫性肺泡损伤, 伴肺泡扩张和塌陷、透明膜形成、间质性水肿、毛细血管充血以及 II 型肺细胞丢失^[14]; 但新冠病毒感染相关 ARDS 患者可见肺部微血栓明显增加^[15], 由于急进性的肺纤维化, 表现出更严重的低氧血症和更高的病死率^[16]。有研究表明, 新冠病毒感染相关 ARDS 患者低氧血症及肺内分流的严重程度常与肺顺应性下降程度不匹配, 这与典型 ARDS 有所不同^[17]。新冠病毒感染相关 ARDS 通常伴随高水平的循环炎症因子, 如肿瘤坏死因子- α (tumor necrosis factor- α , TNF- α) 和白细胞介素-6 (interleukin-6, IL-6), 这可能是导致新冠病毒感染相关 ARDS 的原因^[18]。这种炎症因子风暴不仅限于肺部, 还涉及全身性炎症反应, 包括血管内皮损伤、广泛微血栓形成和肺毛细血管增生^[19-20]。

2.2 线粒体动力学失衡与新冠病毒感染相关 ARDS 的肺功能障碍和损伤: 病毒感染宿主细胞后, 会打破线粒体动力学平衡, 引发线粒体裂变和诱导线粒体自噬异常。例如, 乙型肝炎病毒 (hepatitis B virus, HBV) 抑制线粒体融合, 通过诱导 E3 泛素化连接酶 Parkin 促进线粒体融合蛋白 2 (mitofusin 2, Mfn2) 的泛素化及降解; 另一方面诱导 Drp1 磷酸化促进线粒体裂变, 介导线粒体动力学异常^[21]。流感病毒 PR8 株感染可以导致线粒体片段受损, 诱导线粒体自噬, 加速线粒体碎裂^[22]。委内瑞拉马脑脊髓炎病毒 (Venezuelan equine encephalomyelitis virus, VEEV) 感染的细胞中会出现 Drp1 蛋白富集, 线粒体分裂抑制剂 1 (mitochondrial division inhibitor 1, Mdivi-1) 可以显著降低天冬氨酸特异性半胱氨酸蛋白酶 (caspase) 裂解, 从而支持线粒体裂变是 VEEV 感染细胞凋亡的因素, 提示 VEEV 感染可能也是通过诱发 Drp1 蛋白介导的线粒体裂变促进感染细胞的凋亡^[23]。寨卡病毒 (Zika virus, ZIKV) 的非结构蛋白 NS4A 蛋白过表达后会诱导 Drp1 蛋白的转录后修饰和磷酸化, 引起线粒体裂变, 加重线粒体碎片化^[24]。

新冠病毒的 Spike 蛋白或灭活新冠病毒体外刺激的小胶质细胞呈现出线粒体形态改变以及线粒体的碎片化, 说明新冠病毒也可以引起被感染宿主细胞的动力学失衡^[25-26]。

线粒体动力学失衡会继发线粒体自噬、氧化应激并诱导线粒体 DNA (mitochondrial DNA, mtDNA) 释放,加重肺部损伤和肺功能障碍。新冠病毒感染后引起线粒体动力学失衡,并调控线粒体自噬相关蛋白(如 p62 与 LC3 蛋白)的结合和 Spike 蛋白的表达,使宿主细胞线粒体自噬功能发生损伤,从而促进自身病毒的持续复制和 IL-18 表达的增加及新冠病毒感染相关肺炎炎症反应^[27-28]。新冠病毒相关蛋白可以促进线粒体氧化应激,线粒体活性氧(mitochondrial reactive oxygen species, mtROS)水平升高^[29-31],然而,干扰新冠病毒诱导的线粒体氧化应激可以显著降低内皮功能障碍^[32]。新冠病毒的非结构蛋白 4(nonstructural protein 4, NSP4)和开放阅读框蛋白 9b(open reading frame 9b, ORF9b)诱导形成外膜微孔结构并增加 mtDNA 的释放,mtDNA 进一步活化 NOD 样受体蛋白 3(NOD-like receptor protein 3, NLRP3)炎症小体复合物,介导 caspase-1 依赖的炎症细胞因子释放,诱导线粒体碎片化、钙失衡及细胞凋亡,从而引起线粒体动力学失衡及功能紊乱,这可能是引起新冠病毒感染相关 ARDS 患者“细胞因子风暴”的重要机制^[33-36]。相应的,新冠病毒感染患者血清 mtDNA 含量较健康对照者增加,并激活 Toll 样受体 9(Toll-like receptors 9, TLR9)信号转导,促进炎症反应及内皮功能障碍^[37]。在构建的新冠病毒感染的人血管紧张素转换酶 2(angiotensin-converting enzyme 2, ACE2)转基因小鼠模型中,通过抑制 TLR 诱导的 mtDNA 合成,在体外和体内阻断 NLRP3 炎症小体活化,ARDS 症状显著减轻^[38]。

新冠病毒感染相关 ARDS 中的低氧环境可触发一系列细胞内信号转导,导致缺氧诱导因子-1 α (hypoxia-inducible factor-1 α , HIF-1 α)的表达。新冠病毒的 ORF3a 诱导线粒体损伤和 mtROS 释放,也可以促进 HIF-1 α 的表达^[39-40]。在新冠病毒感染相关 ARDS 患者中, HIF-1 α 可以通过促进新冠病毒的持续感染,触发或增强炎症因子风暴,减少中性粒细胞凋亡,从而造成肺组织局部损伤及多器官损伤的发生,可能是新冠病毒感染相关 ARDS 患者治疗的一个潜在、重要的分子治疗靶点及信号通路^[41]。mtROS/HIF-1 α 信号促进单核细胞和肺泡巨噬细胞的糖酵解过程,而糖酵解也进一步促进新冠病毒持续复制、乳酸释放和炎症因子风暴,导致免疫细胞功能障碍及肺上皮细胞凋亡,促进新冠病毒感染相关 ARDS 的疾病进展^[42-44]。此外, HIF-1 α 还可以通过影响凝血因子和凝血酶原复合物的功能,导致新冠病毒感染相关 ARDS 患者发生凝血级联反应及微血栓形成,最终导致多器官损伤^[45-46]。

3 线粒体动力学平衡与新冠病毒感染相关 ARDS 的治疗

目前新冠病毒感染相关 ARDS 的治疗手段主要包括机械通气(mechanical ventilation, MV)和药物治疗。MV 通过提供足够的氧气和排出二氧化碳,改善患者的氧合状态,减轻呼吸肌疲劳,为患者提供必要的呼吸支持,减少肺损伤,改善肺功能^[47]。在药物治疗方面,糖皮质激素在新冠病毒感染相关 ARDS 治疗中的应用尤为广泛,可以通过抑制炎症反应和减少“细胞因子风暴”来发挥作用^[48]。

3.1 MV 与线粒体动力学平衡:新冠病毒感染相关 ARDS 的病理生理变化导致严重低氧血症和肺顺应性降低,需要 MV 支持,常联合俯卧位通气和体外膜肺氧合(extracorporeal membrane oxygenation, ECMO)治疗^[49-51]。但 MV 本身也可能导致肺损伤及膈肌萎缩和收缩功能障碍,即 MV 相关性肺损伤(ventilation-induced lung injury, VILI)和 MV 相关性膈肌功能障碍(ventilation-induced diaphragmatic dysfunction, VIDD),均是 MV 常见的并发症^[52]。在 MV 的情况下,膈肌和肺组织的线粒体动力学可能因周期性的氧环境变化发生改变。研究表明, MV 6 h 后大鼠横膈膜线粒体外膜的超微亚结构改变,并发生线粒体碎片化, Mfn2 和 OPA1 的表达无变化,而 Drp1 蛋白的表达水平显著增加^[53]。MV 6 h 后,大鼠横膈膜组织的蛋白组学测序结果显示,线粒体功能相关的通路和蛋白存在差异表达,其中 Drp1 蛋白处于上调差异蛋白的前列;电镜结果显示,线粒体结构出现碎片化,线粒体裂变能力增加,而过度的线粒体分裂继发 mtROS 产生和线粒体功能障碍^[54]。同时, MV 诱发的线粒体超分裂会引起线粒体自噬的过度活化和肌浆网钙泄漏,引起膈肌损伤^[55-56]。高潮气量 MV 诱导的机械过度拉伸激活内质网通道蛋白 IP3 受体 1(IP3 receptor 1, IP3R1),而 IP3R 相关的 Ca²⁺ 失调导致内质网应激和线粒体功能障碍,诱导 NLRP3 炎症小体激活,导致 VILI^[57]。MV 可导致线粒体动力学失衡并诱导 HIF-1 α 的表达上调,也可能是参与促进新冠病毒持续复制、炎症因子风暴和多器官损伤的部分机制^[58]。MV 也可导致肺部线粒体功能障碍和肺泡化延迟,通过激活特定的信号通路,导致线粒体功能障碍和肌肉萎缩^[59]。因此,应在临床治疗中采用肺保护性通气策略,如降低潮气量、限制平台压力和使用适当的呼气末正压(positive end-expiratory pressure, PEEP),可以减少 VILI 的发生及对线粒体动力学平衡的不良影响,改善患者的临床结局。

3.2 糖皮质激素与线粒体动力学平衡:糖皮质激素作为治疗重症新冠病毒感染相关 ARDS 的重要药物之一,其作用机制包括抑制炎症反应和调节免疫系统^[60]。糖皮质激素能够抑制炎症细胞(如中性粒细胞、巨噬细胞和淋巴细胞)的活性,并抑制促炎细胞因子(如 TNF 和 IL)的产生,从而减轻炎症因子风暴^[61]。糖皮质激素还可以通过调节免疫细胞的功能,如减少 T 细胞的活化和增殖,进一步控制炎症反应^[62]。而线粒体动力学平衡的调节也可能与糖皮质激素相互作用。糖皮质激素可以通过影响线粒体的分裂和融合过程,间接调节线粒体功能,减轻炎症反应^[63]。糖皮质激素通过影响线粒体内的信号转导途径,如哺乳动物雷帕霉素靶蛋白(mammalian target of rapamycin, mTOR)、AMP 活化蛋白激酶(AMP-activated protein kinase, AMPK)、核转录因子- κ B(nuclear factor- κ B, NF- κ B)和其他炎症信号通路的激活,调节线粒体的代谢状态和自噬过程,进一步减轻线粒体功能障碍,减少炎症介质的产生,从而减轻炎症反应,改善新冠病毒感染相关 ARDS 患者的病情,降低病死率^[48, 64]。糖皮质激素可通过抑制肺组织 NF- κ B 活性,下调巨噬细胞炎症蛋

白-1 α (macrophage inflammatory protein-1 α , MIP-1 α) 的表达,从而减轻肺损伤程度^[65]。糖皮质激素能够直接抑制单核细胞趋化蛋白-1 (monocyte chemoattractant protein-1, MCP-1) 信使 RNA (messenger RNA, mRNA) 的表达,减轻肺泡炎症反应和肺纤维化程度^[66]。糖皮质激素还能够通过影响 TLR 介导的信号转导途径来调节免疫细胞活性。例如,新冠病毒的刺突蛋白可能通过与 TLR4 结合并激活其信号转导,增加细胞表面 ACE2 的表达,从而促进病毒进入宿主细胞^[67]。而糖皮质激素能够抑制 TLR2 和 TLR4 介导的炎症途径,减少炎症细胞因子的产生^[68]。由此可见,糖皮质激素在治疗新冠病毒感染相关 ARDS 中的作用机制是多方面的,包括抑制炎症反应、调节免疫细胞功能以及影响线粒体动力学,并通过调节细胞信号转导途径影响因子释放,改善患者“细胞因子风暴”导致的临床症状,减轻病情严重程度,促进患者恢复。

4 总结与展望

线粒体动力学中分裂与融合的平衡对于维持细胞的正常生理活动起关键作用。新冠病毒感染相关 ARDS 的线粒体动力学失衡,一方面,由于严重的低氧血症引起的 Drp1 介导线粒体超分裂;另一方面,新冠病毒的蛋白成分也与线粒体相互作用,介导线粒体功能紊乱,并引起肺损伤和肺功能障碍。

目前,现有的治疗手段仍存在局限性。MV 可能加重肺损伤及膈肌损伤,导致 VILI 及 VIDD,不恰当的 MV 策略也可能导致高病死率。糖皮质激素可能会引起免疫抑制 (immunosuppression, IS) 作用,延缓病毒清除,以及导致神经系统和代谢紊乱、继发感染等不良反应的发生。未来的研究可侧重于通过分子生物学和细胞生物学方法,明确线粒体功能障碍如何影响新冠病毒感染的病理过程,以及如何通过调节线粒体功能来减轻病毒引起的肺损伤,并基于线粒体在新冠病毒感染相关 ARDS 中的作用,开发新的治疗药物或疗法,如线粒体靶向抗氧化剂、调节线粒体通透性孔蛋白活性的药物等,以减轻炎症反应和氧化应激,保护肺组织。研究现有的抗病毒、抗炎和免疫调节治疗策略对线粒体功能的影响,以及这些治疗策略是否能够通过改善线粒体功能来提高治疗效果,以期对新冠病毒感染相关 ARDS 提供新的治疗思路。虽然线粒体动力学和线粒体自噬有望成为 IS 的新靶点,但是如何实现线粒体动力学中分裂与融合的平衡以及如何发挥线粒体自噬“双刃剑”中“利”的一面,仍需更加深入地探索研究。

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参考文献

- [1] Wang DW, Hu B, Hu C, et al. Clinical characteristics of 138 hospitalized patients with 2019 novel coronavirus-infected pneumonia in Wuhan, China [J]. *JAMA*, 2020, 323 (11): 1061-1069. DOI: 10.1001/jama.2020.1585. Erratum in: *JAMA*, 2021, 325 (11): 1113. DOI: 10.1001/jama.2021.2336.
- [2] Dutra Silva J, Su Y, Calfee CS, et al. Mesenchymal stromal cell extracellular vesicles rescue mitochondrial dysfunction and improve barrier integrity in clinically relevant models of ARDS [J]. *Eur Respir J*, 2021, 58 (1): 2002978. DOI: 10.1183/13993003.02978-2020.
- [3] Matthay MA, Arabi YM, Siegel ER, et al. Phenotypes and personalized medicine in the acute respiratory distress syndrome [J]. *Intensive Care Med*, 2020, 46 (12): 2136-2152. DOI: 10.1007/s00134-020-06296-9.
- [4] Tilokani L, Nagashima S, Paupe V, et al. Mitochondrial dynamics: overview of molecular mechanisms [J]. *Essays Biochem*, 2018, 62 (3): 341-360. DOI: 10.1042/EBBC20170104.
- [5] Chen W, Zhao HK, Li YS. Mitochondrial dynamics in health and disease: mechanisms and potential targets [J]. *Signal Transduct Target Ther*, 2023, 8 (1): 333. DOI: 10.1038/s41392-023-01547-9.
- [6] Adebayo M, Singh S, Singh AP, et al. Mitochondrial fusion and fission: the fine-tune balance for cellular homeostasis [J]. *FASEB J*, 2021, 35 (6): e21620. DOI: 10.1096/fj.202100067R.
- [7] Scheffer DDL, Garcia AA, Lee L, et al. Mitochondrial fusion, fission, and mitophagy in cardiac diseases: challenges and therapeutic opportunities [J]. *Antioxid Redox Signal*, 2022, 36 (13-15): 844-863. DOI: 10.1089/ars.2021.0145.
- [8] Jiang J, Wang J, Li C, et al. Hyperoxia induces alveolar epithelial cell apoptosis by regulating mitochondrial function through small mothers against decapentaplegic 3 (SMAD3) and extracellular signal-regulated kinase 1/2 (ERK1/2) [J]. *Bioengineered*, 2022, 13 (1): 242-252. DOI: 10.1080/21655979.2021.2012953.
- [9] Mao CP, Zhang JJ, Lin SC, et al. MiRNA-30a inhibits AECs-II apoptosis by blocking mitochondrial fission dependent on Drp-1 [J]. *J Cell Mol Med*, 2014, 18 (12): 2404-2416. DOI: 10.1111/jcmm.12420.
- [10] 俞典, 谢永鹏, 李小民. 基于 Hippo 信号通路探究自噬在脓毒症 ARDS 肺损伤中的作用机制 [J]. *中华危重病急救医学*, 2023, 35 (8): 884-888. DOI: 10.3760/cma.j.cn121430-20220705-00633.
- [11] Zheng XL, Qian YB, Fu BQ, et al. Mitochondrial fragmentation limits NK cell-based tumor immunosurveillance [J]. *Nat Immunol*, 2019, 20 (12): 1656-1667. DOI: 10.1038/s41590-019-0511-1.
- [12] Quintana-Cabrera R, Scorrano L. Determinants and outcomes of mitochondrial dynamics [J]. *Mol Cell*, 2023, 83 (6): 857-876. DOI: 10.1016/j.molcel.2023.02.012.
- [13] Long Y, Ang Y, Chen W, et al. Hydrogen alleviates impaired lung epithelial barrier in acute respiratory distress syndrome via inhibiting Drp1-mediated mitochondrial fission through the Trx1 pathway [J]. *Free Radic Biol Med*, 2024, 218: 132-148. DOI: 10.1016/j.freeradbiomed.2024.03.022.
- [14] Carsana L, Sonzogni A, Nasr A, et al. Pulmonary post-mortem findings in a series of COVID-19 cases from northern Italy: a two-centre descriptive study [J]. *Lancet Infect Dis*, 2020, 20 (10): 1135-1140. DOI: 10.1016/S1473-3099(20)30434-5.
- [15] Hariji LP, North CM, Shih AR, et al. Lung histopathology in coronavirus disease 2019 as compared with severe acute respiratory syndrome and H1N1 influenza: a systematic review [J]. *Chest*, 2021, 159 (1): 73-84. DOI: 10.1016/j.chest.2020.09.259.
- [16] Wu CM, Chen XY, Cai YP, et al. Risk factors associated with acute respiratory distress syndrome and death in patients with coronavirus disease 2019 pneumonia in Wuhan, China [J]. *JAMA Intern Med*, 2020, 180 (7): 934-943. DOI: 10.1001/jamainternmed.2020.0994. Erratum in: *JAMA Intern Med*, 2020, 180 (7): 1031. DOI: 10.1001/jamainternmed.2020.1429.
- [17] Gattinoni L, Coppola S, Cressoni M, et al. COVID-19 does not lead to a "typical" acute respiratory distress syndrome [J]. *Am J Respir Crit Care Med*, 2020, 201 (10): 1299-1300. DOI: 10.1164/rccm.202003-0817LE.
- [18] Mattos Pereira V, Pradhanang S, Thakar A, et al. Targeting ADAM-17 to alleviate cytokine storm associated with COVID-19 [J]. *FASEB J*, 2022, 36.
- [19] Ackermann M, Mentzer SJ, Kolb M, et al. Inflammation and intussusceptive angiogenesis in COVID-19: everything in and out of flow [J]. *Eur Respir J*, 2020, 56 (5): 2003147. DOI: 10.1183/13993003.03147-2020.
- [20] Ackermann M, Verleden SE, Kuehnel M, et al. Pulmonary vascular endothelialitis, thrombosis, and angiogenesis in Covid-19 [J]. *N Engl J Med*, 2020, 383 (2): 120-128. DOI: 10.1056/NEJMoA2015432.
- [21] Kim SJ, Khan M, Quan J, et al. Hepatitis B virus disrupts mitochondrial dynamics: induces fission and mitophagy to attenuate apoptosis [J]. *PLoS Pathog*, 2013, 9 (12): e1003722. DOI: 10.1371/journal.ppat.1003722.
- [22] Lee JH, Oh SJ, Yun J, et al. Nonstructural protein NS1 of influenza virus disrupts mitochondrial dynamics and enhances mitophagy via ULK1 and BNIP3 [J]. *Viruses*, 2021, 13 (9): 1845. DOI: 10.3390/v13091845.
- [23] Keck F, Brooks-Faulconer T, Lark T, et al. Altered mitochondrial dynamics as a consequence of Venezuelan equine encephalitis virus infection [J]. *Virulence*, 2017, 8 (8): 1849-1866. DOI: 10.1080/21505594.2016.1276690.
- [24] Lee JK, Shin OS. Zika virus modulates mitochondrial dynamics, mitophagy, and mitochondria-derived vesicles to facilitate viral replication in trophoblast cells [J]. *Front Immunol*, 2023, 14: 1203645. DOI: 10.3389/fimmu.2023.1203645.
- [25] Clough E, Inigo J, Chandra D, et al. Mitochondrial dynamics in SARS-CoV2 spike protein treated human microglia: implications for neuro-COVID [J]. *J Neuroimmune Pharmacol*, 2021, 16 (4): 770-784.

- DOI: 10.1007/s11481-021-10015-6. Erratum in: J Neuroimmune Pharmacol, 2022, 17 (3-4): 380. DOI: 10.1007/s11481-021-10043-2.
- [26] Cortese M, Lee JY, Cerikan B, et al. Integrative imaging reveals SARS-CoV-2-induced reshaping of subcellular morphologies [J]. Cell Host Microbe, 2020, 28 (6): 853-866. e5. DOI: 10.1016/j.chom.2020.11.003.
- [27] Shang C, Liu ZR, Zhu YL, et al. SARS-CoV-2 causes mitochondrial dysfunction and mitophagy impairment [J]. Front Microbiol, 2022, 12: 780768. DOI: 10.3389/fmicb.2021.780768.
- [28] Liang SX, Bao CL, Yang Z, et al. SARS-CoV-2 spike protein induces IL-18-mediated cardiopulmonary inflammation via reduced mitophagy [J]. Signal Transduct Target Ther, 2023, 8 (1): 108. DOI: 10.1038/s41392-023-01368-w.
- [29] Zhang YH, Peng XY, Xue MJ, et al. SARS-CoV-2 spike protein promotes RPE cell senescence via the ROS/P53/P21 pathway [J]. Biogerontology, 2023, 24 (5): 813-827. DOI: 10.1007/s10522-023-10019-0.
- [30] Yu HY, Yang L, Han ZN, et al. SARS-CoV-2 nucleocapsid protein enhances the level of mitochondrial reactive oxygen species [J]. J Med Virol, 2023, 95 (12): e29270. DOI: 10.1002/jmv.29270.
- [31] Guarnieri JW, Lie T, Albrecht YES, et al. Mitochondrial antioxidants abate SARS-CoV-2 pathology in mice [J]. Proc Natl Acad Sci U S A, 2024, 121 (30): e2321972121. DOI: 10.1073/pnas.2321972121.
- [32] Youn JY, Wang J, Li Q, et al. Robust therapeutic effects on COVID-19 of novel small molecules: alleviation of SARS-CoV-2 S protein induction of ACE2/TMPRSS2, NOX2/ROS, and MCP-1 [J]. Front Cardiovasc Med, 2022, 9: 957340. DOI: 10.3389/fcvm.2022.957340.
- [33] Mahmoodpoor A, Sanaie S, Ostadi Z, et al. Roles of mitochondrial DNA in dynamics of the immune response to COVID-19 [J]. Gene, 2022, 836: 146681. DOI: 10.1016/j.gene.2022.146681.
- [34] Gatti P, Ilamathi HS, Todkar K, et al. Mitochondria targeted viral replication and survival strategies—prospective on SARS-CoV-2 [J]. Front Pharmacol, 2020, 11: 578599. DOI: 10.3389/fphar.2020.578599.
- [35] Faizan MI, Chaudhuri R, Sagar S, et al. NSP4 and ORF9b of SARS-CoV-2 induce pro-inflammatory mitochondrial DNA release in inner membrane-derived vesicles [J]. Cells, 2022, 11 (19): 2969. DOI: 10.3390/cells11192969.
- [36] Guarnieri JW, Angelin A, Murdock DG, et al. SARS-CoV-2 viroporins activate the NLRP3-inflammasome by the mitochondrial permeability transition pore [J]. Front Immunol, 2023, 14: 1064293. DOI: 10.3389/fimmu.2023.1064293.
- [37] Costa TJ, Potje SR, Fraga-Silva TFC, et al. Mitochondrial DNA and TLR9 activation contribute to SARS-CoV-2-induced endothelial cell damage [J]. Vascul Pharmacol, 2022, 142: 106946. DOI: 10.1016/j.vph.2021.106946.
- [38] Xian HX, Liu Y, Rundberg Nilsson A, et al. Metformin inhibition of mitochondrial ATP and DNA synthesis abrogates NLRP3 inflammasome activation and pulmonary inflammation [J]. Immunity, 2021, 54 (7): 1463-1477. e11. DOI: 10.1016/j.immuni.2021.05.004.
- [39] Tian MF, Liu WY, Li X, et al. HIF-1 α promotes SARS-CoV-2 infection and aggravates inflammatory responses to COVID-19 [J]. Signal Transduct Target Ther, 2021, 6 (1): 308. DOI: 10.1038/s41392-021-00726-w.
- [40] Jana S, Heaven MR, Stauffi CB, et al. HIF-1 α -dependent metabolic reprogramming, oxidative stress, and bioenergetic dysfunction in SARS-CoV-2-infected hamsters [J]. Int J Mol Sci, 2022, 24 (1): 558. DOI: 10.3390/ijms24010558.
- [41] Serebrovska ZO, Chong EY, Serebrovska TV, et al. Hypoxia, HIF-1 α , and COVID-19: from pathogenic factors to potential therapeutic targets [J]. Acta Pharmacol Sin, 2020, 41 (12): 1539-1546. DOI: 10.1038/s41401-020-00554-8.
- [42] Codo AC, Davanzo GG, Monteiro LB, et al. Elevated glucose levels favor SARS-CoV-2 infection and monocyte response through a HIF-1 α /glycolysis-dependent axis [J]. Cell Metab, 2020, 32 (3): 437-446. e5. DOI: 10.1016/j.cmet.2020.07.007. Erratum in: Cell Metab, 2020, 32 (3): 498-499. DOI: 10.1016/j.cmet.2020.07.015.
- [43] Ferraro E, Germanò M, Mollace R, et al. HIF-1, the warburg effect, and macrophage/microglia polarization potential role in COVID-19 pathogenesis [J]. Oxid Med Cell Longev, 2021, 2021: 8841911. DOI: 10.1155/2021/8841911.
- [44] Zhu BB, Wu Y, Huang S, et al. Uncoupling of macrophage inflammation from self-renewal modulates host recovery from respiratory viral infection [J]. Immunity, 2021, 54 (6): 1200-1218. e9. DOI: 10.1016/j.immuni.2021.04.001.
- [45] Zalpoor H, Akbari A, Nabi-Afjadi M, et al. Hypoxia-inducible factor 1 alpha (HIF-1 α) stimulated and P2X7 receptor activated by COVID-19, as a potential therapeutic target and risk factor for epilepsy [J]. Hum Cell, 2022, 35 (5): 1338-1345. DOI: 10.1007/s13577-022-00747-9.
- [46] Ambade V, Ambade S. SARS-CoV-2 infecting endothelial cells, biochemical alterations, autopsy findings and outcomes in COVID-19, suggest role of hypoxia-inducible factor-1 [J]. J Med Biochem, 2022, 41 (1): 14-20. DOI: 10.5937/jomb0-30659.
- [47] Kondili E, Makris D, Georgopoulos D, et al. COVID-19 ARDS: points to be considered in mechanical ventilation and weaning [J]. J Pers Med, 2021, 11 (11): 1109. DOI: 10.3390/jpm11111109.
- [48] Feng YA, Li HC. Further discussion on glucocorticoid treatment of COVID-19 [J]. Chin Med J (Engl), 2022, 135 (11): 1386. DOI: 10.1097/CM9.0000000000001872.
- [49] Akyar İ. COVID-19 hastalarında akut solunum sıkıntısı sendromu (ARDS) yönetimi ve hemşirelik bakımı [J]. Hacettepe Univ Fac Nurs, 2020, 7: 8-14.
- [50] 徐燕, 孟玫, 刘娇, 等. 危重型新型冠状病毒肺炎患者俯卧位通气实操流程 [J]. 中华危重病急救医学, 2021, 33 (4): 393-398. DOI: 10.3760/cma.j.cn121430-20210224-00112.
- [51] 焦国庆, 姜淑云, 陈静瑜, 等. 联合体外膜肺氧合在新型冠状病毒肺炎肺移植术中的应用体会 [J]. 中华危重病急救医学, 2021, 33 (12): 1504-1507. DOI: 10.3760/cma.j.cn121430-20210130-00181.
- [52] Kondili E, Makris D, Georgopoulos D, et al. COVID-19 ARDS: points to be considered in mechanical ventilation and weaning [J]. J Pers Med, 2021, 11 (11): 1109. DOI: 10.3390/jpm11111109.
- [53] Picard M, Azuelos I, Jung B, et al. Mechanical ventilation triggers abnormal mitochondrial dynamics and morphology in the diaphragm [J]. J Appl Physiol (1985), 2015, 118 (9): 1161-1171. DOI: 10.1152/japplphysiol.00873.2014.
- [54] Dridi H, Yehya M, Barsotti R, et al. Aberrant mitochondrial dynamics contributes to diaphragmatic weakness induced by mechanical ventilation [J]. PNAS Nexus, 2023, 2 (11): 336. DOI: 10.1093/pnasnexus/pgad336.
- [55] Yong H, Zhou Y, Ye WL, et al. PINK1/Parkin-mediated mitophagy in mechanical ventilation-induced diaphragmatic dysfunction [J]. Ther Adv Respir Dis, 2021, 15: 1753466621998246. DOI: 10.1177/1753466621998246.
- [56] Dridi H, Yehya M, Barsotti R, et al. Mitochondrial oxidative stress induces leaky ryanodine receptor during mechanical ventilation [J]. Free Radic Biol Med, 2020, 146: 383-391. DOI: 10.1016/j.freeradbiomed.2019.11.019.
- [57] Ye L, Zeng Q, Ling MY, et al. Inhibition of IP3R/Ca²⁺ dysregulation protects mice from ventilator-induced lung injury via endoplasmic reticulum and mitochondrial pathways [J]. Front Immunol, 2021, 12: 729094. DOI: 10.3389/fimmu.2021.729094.
- [58] Li LF, Yu CC, Wu HP, et al. Reduction in ventilation-induced diaphragmatic mitochondrial injury through hypoxia-inducible factor 1 α in a murine endotoxemia model [J]. Int J Mol Sci, 2022, 23 (3): 1083. DOI: 10.3390/ijms23031083.
- [59] Nguyen Thi Dieu T, Pham Nhat A, Craig TJ, et al. Clinical characteristics and cytokine changes in children with pneumonia requiring mechanical ventilation [J]. J Int Med Res, 2017, 45 (6): 1805-1817. DOI: 10.1177/0300060516672766.
- [60] Jin SQ, Ding XB, Yang CX, et al. Mechanical ventilation exacerbates Poly (I:C) induced acute lung injury: central role for caspase-11 and gut-lung axis [J]. Front Immunol, 2021, 12: 693874. DOI: 10.3389/fimmu.2021.693874.
- [61] Prescott HC, Rice TW. Corticosteroids in COVID-19 ARDS: evidence and hope during the pandemic [J]. JAMA, 2020, 324 (13): 1292-1295. DOI: 10.1001/jama.2020.16747.
- [62] Andreaskos E, Papadakis M, Serhan CN. Dexamethasone, pro-resolving lipid mediators and resolution of inflammation in COVID-19 [J]. Allergy, 2021, 76 (3): 626-628. DOI: 10.1111/all.14595.
- [63] Qin C, Zhou LQ, Hu ZW, et al. Dysregulation of immune response in patients with coronavirus 2019 (COVID-19) in Wuhan, China [J]. Clin Infect Dis, 2020, 71 (15): 762-768. DOI: 10.1093/cid/ciaa248.
- [64] Lu M, Guo JD, Wu BW, et al. Mesenchymal stem cell-mediated mitochondrial transfer: a therapeutic approach for ischemic stroke [J]. Transl Stroke Res, 2021, 12 (2): 212-229. DOI: 10.1007/s12975-020-00853-6.
- [65] Leach DA, Brooke GN, Bevan CL. Roles of steroid receptors in the lung and COVID-19 [J]. Essays Biochem, 2021, 65 (6): 1025-1038. DOI: 10.1042/EBC20210005.
- [66] Liu ZH, Zhang XR, Hu XY, et al. Effect of glucocorticoid on MIP-1 α and NF- κ B expressing in the lung of rats undergoing mechanical ventilation with a high tidal volume [J]. World J Emerg Med, 2011, 2 (1): 66-69.
- [67] Aboudounya MM, Heads RJ. COVID-19 and Toll-like receptor 4 (TLR4): SARS-CoV-2 may bind and activate TLR4 to increase ACE2 expression, facilitating entry and causing hyperinflammation [J]. Mediators Inflamm, 2021, 2021: 8874339. DOI: 10.1155/2021/8874339.
- [68] Sævik AB, Ueland G, Åkerman AK, et al. Altered biomarkers for cardiovascular disease and inflammation in autoimmune Addison's disease – a cross-sectional study [J]. Eur J Endocrinol, 2023, 189 (4): 438-447. DOI: 10.1093/ejendo/lvad136.