

# 嗜铬粒蛋白 A 衍生多肽 CGA<sub>47-66</sub> 抑制脓毒症小鼠血脑屏障通透性增加

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**【摘要】** 目的 探讨嗜铬粒蛋白 A (CGA) 衍生多肽 CGA<sub>47-66</sub> (CHR) 对脓毒症小鼠血脑屏障通透性的影响。方法 健康雄性 C57BL/6 小鼠 120 只,按随机数字表法分组,每组 12 只。72 只用于观察脂多糖 (LPS) 诱导小鼠脑组织含水量和脑组织伊文思蓝 (EB) 含量的动态变化。另 48 只分为生理盐水对照组 (NS 组)、LPS 诱导脓毒症模型组 (LPS 组)、低剂量 CHR 预处理组 (C<sub>L</sub>+LPS 组) 及高剂量 CHR 预处理组 (C<sub>H</sub>+LPS 组)。腹腔注射 10 mg/kg LPS 0.1 mL 制备脓毒症小鼠模型; NS 组注射等量生理盐水; C<sub>L</sub>+LPS 组和 C<sub>H</sub>+LPS 组于注射 LPS 前 10 min 分别腹腔注射 CHR 15.5 μg/kg 和 77.5 μg/kg。6 h 后经尾静脉注射 2% EB 4 mL/kg,检测脑组织含水量 and EB 含量;采用荧光示踪法定性观察血脑屏障通透性变化;苏木素-伊红 (HE) 染色后观察脑组织病理学改变。结果 注射 LPS 后随着时间的延长,脑组织含水量和 EB 含量均逐渐增加,脑组织含水量与对照组出现统计学差异的时间早于 EB 含量变化的时间 (分别为 3 h 和 6 h)。LPS 组脑组织含水量及 EB 含量均较 NS 组明显升高 [脑组织含水量: (79.77±0.62)% 比 (78.28±0.44)%,  $P<0.01$ ; EB 含量 (μg/g): 13.87±4.50 比 7.13±1.76,  $P<0.05$ ]; 而两个 CHR 预处理组均可逆转 LPS 诱导的脑组织含水量和 EB 含量增高,以 C<sub>H</sub>+LPS 组效果更加明显 [脑组织含水量: (78.15±0.73)% 比 (79.77±0.62)%, EB (μg/g): 7.09±2.59 比 13.87±4.50, 均  $P<0.05$ ]。EB 荧光观察结果显示, NS 组荧光信号仅存在于脑膜; LPS 组脑实质中可见 EB 荧光广泛分布,说明 EB 渗漏较 NS 组明显;两个 CHR 预处理组脑实质中 EB 荧光减少,说明 EB 渗漏明显减少,且以高剂量组改善更为明显。HE 染色显示, NS 组脑血管结构清晰,周围间隙未见明显增大; LPS 组脑组织结构疏松,小血管周围间隙明显增宽,水肿明显;两个 CHR 预处理组脑水肿较 LPS 组有所改善,以高剂量组作用更加明显。结论 LPS 诱导脓毒症小鼠血脑屏障通透性变化呈时间依赖性;使用外源性 CGA 衍生多肽 CHR 可抑制 LPS 诱导脓毒症小鼠血脑屏障通透性增加,减轻脑水肿,保护脑组织,以 77.5 μg/kg 高剂量 CHR 效果更加明显。

**【关键词】** 脓毒症; 嗜铬粒蛋白 A; 嗜铬粒蛋白 A 衍生多肽; 血脑屏障

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**Chromogranin A derived peptide CGA<sub>47-66</sub> inhibits hyper-permeability of blood brain barrier in mice with sepsis** Zeng Yan, Zhang Dan, Jiang Liping, Wei Fu, Xu Shan

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**【Abstract】 Objective** To explore the effect of chromofungin (CHR), a chromogranin A (CGA) derived peptide CGA<sub>47-66</sub>, on hyper-permeability of blood brain barrier in septic mice. **Methods** 120 healthy male C57BL/6 mice were randomly divided into groups, with 12 mice in each group. Seventy-two mice were used for dynamic observation of the contents of water and Evan blue (EB) in brain tissue after being treated with lipopolysaccharide (LPS). Another 48 mice were divided into normal saline control group (NS group), LPS induced sepsis model group (LPS group), low-dose CHR pretreatment group (C<sub>L</sub>+LPS group), and high-dose CHR pretreatment group (C<sub>H</sub>+LPS group). The septic model was reproduced by intraperitoneal injection of 10 mg/kg LPS 0.1 mL, and the mice in NS group was given equal volume of normal saline. The mice in C<sub>L</sub>+LPS group and C<sub>H</sub>+LPS group were intraperitoneally injected with 15.5 μg/kg and 77.5 μg/kg CHR 10 minutes before LPS injection. Six hours after LPS injection, 4 mL/kg of 2% EB was injected via caudal vein, the contents of water and EB in brain tissue were determined, and EB immune fluorescence in brain tissue was determined to assess the changes in permeability of blood brain barrier. Brain pathology was observed with hematoxylin and eosin (HE) staining. **Results** With the extension of time after LPS injection, the contents of water and EB in brain tissue were gradually increased, and the time of difference with statistical significance appeared earlier when compared with

that of control group in the contents of water than that in EB contents (3 hours and 6 hours, respectively). The contents of water and EB in brain tissue in LPS group were significantly increased as compared with NS group [water content:  $(79.77 \pm 0.62)\%$  vs.  $(78.28 \pm 0.44)\%$ ,  $P < 0.01$ ; EB content ( $\mu\text{g/g}$ ):  $13.87 \pm 4.50$  vs.  $7.13 \pm 1.76$ ,  $P < 0.05$ ]. CHR pretreatment with either of two dosages could reverse the increase in water and EB contents in brain tissue induced by LPS, and the effect was more significant in  $C_H$ +LPS group [water content:  $(78.15 \pm 0.73)\%$  vs.  $(79.77 \pm 0.62)\%$ , EB ( $\mu\text{g/g}$ ):  $7.09 \pm 2.59$  vs.  $13.87 \pm 4.50$ , both  $P < 0.05$ ]. It was shown by EB fluorescence observation that the fluorescence signal displayed only in the meninges in NS group, and EB fluorescence was widely distributed in brain parenchyma in LPS group, indicating that the EB leakage in LPS group was more marked than that of NS group. In CHR pretreatment groups, EB fluorescence was decreased in brain parenchyma, indicating that EB leakage was significantly less marked, while it was more obvious in high dose CHR group. It was shown by HE staining that cerebral blood vessel structure was intact in NS group, and the gap around blood vessel was not significant increased. On the other hand, brain structure in LPS group appeared loose, with widening of small perivascular spaces and obvious edema. Brain edema in CHR pretreatment groups was improved as compared with that of the LPS group, and it was more apparent in high dose CHR group.

**Conclusions** LPS induced change in blood brain barrier permeability in mice in a time-dependent manner. Exogenous CGA derived peptides CHR can inhibit LPS induced hyper-permeability of blood brain barrier in septic mice, thus reduces brain edema, protects the brain tissue, and the effect is more obvious with a high dose of CHR ( $77.5 \mu\text{g/kg}$ ).

**【Key words】** Sepsis; Chromogranin A; Chromogranin A derived peptide; Blood brain barrier

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脓毒症是由感染引起的全身炎症反应失控导致的多器官功能障碍<sup>[1-3]</sup>,脓毒症相关性脑病(SAE)是由脓毒症引起的脑功能损害,常表现为意识和认知功能障碍,是脓毒症常见并发症之一<sup>[4]</sup>。研究证实,SAE可显著增加脓毒症患者病死率<sup>[5]</sup>,但由于其发病机制复杂<sup>[6-9]</sup>,尚未完全阐明,且目前尚无有效的针对性治疗方法。现有研究表明,全身炎症反应可引起血脑屏障功能损害<sup>[10-11]</sup>,可能是SAE发病机制的重要环节,因此,改善血脑屏障功能障碍可能是治疗SAE的重要手段。嗜铬粒蛋白A(CGA)是粒蛋白家族的重要成员,在肾上腺髓质中,嗜铬粒细胞与儿茶酚胺共同释放,在机体炎症应激中发挥作用。血脑屏障功能正常与脑微血管内皮细胞稳态密切相关。本课题组前期体外研究发现,CGA的NH<sub>2</sub>端多肽CGA<sub>47-66</sub>(即CHR)可以抑制脓毒症患者血清或肿瘤坏死因子- $\alpha$ (TNF- $\alpha$ )引起的血管内皮骨架蛋白重组,减少钙黏蛋白表达,增加血管内皮通透性<sup>[12]</sup>;而有关CHR在体内是否能改善血脑屏障通透性的研究尚未见报道。我们假设CHR在体内也能作用于脑微血管内皮,改善血脑屏障通透性,进而改善SAE,故设计本研究,通过腹腔注射脂多糖(LPS)制备脓毒症小鼠模型,探讨CGA衍生多肽CHR对其血脑屏障通透性的调节作用。

## 1 材料与方法

**1.1 实验动物及材料:** SPF级健康雄性C57BL/6小鼠120只,体质量18~22g,6~8周龄,由重庆医科

大学实验动物中心提供,动物合格证号:SCXK(渝)2012-0002。CHR购自上海科肽生物科技有限公司,LPS、伊文思蓝(EB)购自美国Sigma公司。

**1.2 研究分组及处理:**取72只小鼠,按随机数字表法分为LPS作用1、3、6、12、24h组和对照组,每组12只。LPS各组腹腔注射10mg/kg LPS 0.1mL,对照组给予等量生理盐水。观察两组小鼠脑含水量及EB含量的动态变化。另取48只小鼠按随机数字表法分为生理盐水对照组(NS组)、LPS组、低剂量CHR预处理组( $C_L$ +LPS组)及高剂量CHR预处理组( $C_H$ +LPS组),每组12只,6只用于测定脑含水量,6只用于测定脑组织EB含量和组织学观察。LPS组腹腔注射10mg/kg LPS 0.1mL;NS组注射等量生理盐水; $C_L$ +LPS组和 $C_H$ +LPS组于注射LPS前10min分别腹腔注射15.5 $\mu\text{g/kg}$ 、77.5 $\mu\text{g/kg}$  CHR。

**1.3 伦理学:**本实验动物处置方法符合动物伦理学标准。

## 1.4 检测指标及方法

**1.4.1 脑组织含水量测定:**断颈处死各组6只小鼠取大脑皮质及海马组织100mg,称湿质量,烤箱中烘干48h后称干质量,计算脑组织含水量[(湿质量-干质量)/湿质量 $\times 100\%$ ]。

**1.4.2 脑组织EB含量和荧光测定及病理学观察:**各组另取6只小鼠经尾静脉注射2%EB 4mL/kg,腹腔麻醉后暴露小鼠胸腔,穿刺左心室,剪破右心房,以20mL生理盐水灌注直至流出清亮无色

液体。处死小鼠后开颅取皮质及海马区域脑组织。

① 取部分脑组织后置于二甲基甲酰胺溶液中 60℃ 水浴 24 h,离心 10 min 取上清,在波长 632 nm 处测定吸光度(A)值,根据 EB 标准曲线计算脑组织 EB 含量,以  $\mu\text{g/g}$  表示。② 取部分脑组织立即置入液氮中冷冻保存,连续冠状切片,片厚 30  $\mu\text{m}$ ,直接贴于载玻片上,在奥林巴斯 BX60 倒置荧光显微镜 G 激发状态下 EB 呈红色荧光,观察 EB 荧光位置及强度。③ 取部分脑组织用 4% 多聚甲醛固定 24 h,洗去固定液,浓度梯度乙醇脱水,二甲苯透明后浸蜡包埋、切片,并进行苏木素-伊红(HE)染色,观察脑组织病理学变化。

**1.5 统计学处理:**应用 SPSS 19.0 统计软件,计量资料以均数  $\pm$  标准差( $\bar{x} \pm s$ )表示,组间比较采用方差分析(ANOVA),两两比较采用 *t* 检验; $P < 0.05$  为差异有统计学意义。

**2 结果**

**2.1 脑组织含水量及 EB 含量动态变化(表 1):**与对照组相比,LPS 注射后 3 h 和 6 h 小鼠脑组织含水量明显升高(均  $P < 0.01$ );随后脑组织含水量呈逐渐下降趋势,并接近对照组水平。LPS 注射后 6 h 起小鼠脑组织 EB 含量较对照组明显升高,并持续至 24 h(均  $P < 0.01$ )。说明在脓毒症早期,血脑屏障通透性已发生改变,且时间越长,血脑屏障通透性变化越明显。

组别	动物数(只)	脑含水量(%)	EB 含量( $\mu\text{g/g}$ )
对照组	12	78.24 $\pm$ 0.50	7.75 $\pm$ 1.23
LPS 1 h 组	12	79.25 $\pm$ 1.35	6.55 $\pm$ 2.32
LPS 3 h 组	12	80.75 $\pm$ 1.27 <sup>a</sup>	6.96 $\pm$ 1.94
LPS 6 h 组	12	80.14 $\pm$ 0.49 <sup>a</sup>	19.39 $\pm$ 3.09 <sup>a</sup>
LPS 12 h 组	12	78.31 $\pm$ 0.27	20.53 $\pm$ 6.45 <sup>a</sup>
LPS 24 h 组	12	78.40 $\pm$ 0.30	31.79 $\pm$ 4.61 <sup>a</sup>
<i>F</i> 值		8.917	67.763
<i>P</i> 值		0.000	0.000

注:LPS 为脂多糖,EB 为伊文思蓝;与对照组比较,<sup>a</sup> $P < 0.01$

**2.2 CHR 预处理对脑组织含水量及 EB 含量的影响(表 2):**LPS 组脑组织含水量和 EB 含量均较 NS 组明显升高( $P < 0.01$  和  $P < 0.05$ );不同剂量 CHR 预处理组脑组织含水量和 EB 含量均较 LPS 组下降,但仅  $C_H$ +LPS 组表现出统计学意义(均  $P < 0.05$ )。

表 2 CHR 对 LPS 诱导脓毒症小鼠脑含水量及 EB 含量的影响( $\bar{x} \pm s$ )

组别	动物数(只)	脑含水量(%)	EB 含量( $\mu\text{g/g}$ )
NS 组	12	78.28 $\pm$ 0.44	7.13 $\pm$ 1.76
LPS 组	12	79.77 $\pm$ 0.62 <sup>a</sup>	13.87 $\pm$ 4.50 <sup>b</sup>
$C_L$ +LPS 组	12	79.46 $\pm$ 0.88	11.13 $\pm$ 2.71
$C_H$ +LPS 组	12	78.15 $\pm$ 0.73 <sup>c</sup>	7.09 $\pm$ 2.59 <sup>c</sup>
<i>F</i> 值		10.301	6.632
<i>P</i> 值		0.000	0.003

注:CHR 为嗜铬粒蛋白 A 衍生多肽 CGA<sub>47-66</sub>,LPS 为脂多糖,EB 为伊文思蓝,NS 为生理盐水, $C_L$  为 15.5  $\mu\text{g/kg}$  CHR, $C_H$  为 77.5  $\mu\text{g/kg}$  CHR;与 NS 组比较,<sup>a</sup> $P < 0.01$ ,<sup>b</sup> $P < 0.05$ ;与 LPS 组比较,<sup>c</sup> $P < 0.05$

**2.3 脑组织 EB 荧光观察结果:**荧光显微镜下显示,NS 组小鼠脑膜呈线形红色荧光,脑实质内几乎无明显荧光信号(图 1A);LPS 组大脑全脑呈广泛红色荧光斑,以毛细血管为中心向周围弥散(图 1B); $C_L$ +LPS 组和  $C_H$ +LPS 组红色荧光较 LPS 组有所减少,以  $C_H$ +LPS 组减少更明显(图 1C ~ 1D)。

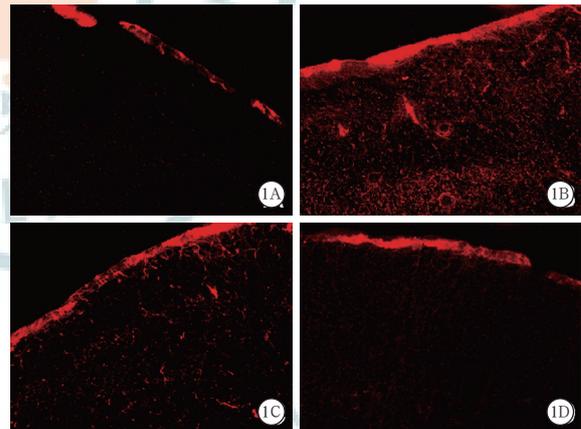


图 1 荧光显微镜下观察嗜铬粒蛋白 A 衍生多肽 CGA<sub>47-66</sub> (CHR)对脂多糖(LPS)诱导脓毒症小鼠脑皮质伊文思蓝(EB)渗漏的影响 EB 在 G 激发状态下呈红色荧光。生理盐水对照组(NS 组, A)脑膜见明显红色荧光,脑实质中几乎见不到荧光;LPS 组(B)除脑膜可见 EB 线形荧光外,脑实质中红色荧光分布广泛,以脑血管为中心向周围弥散,EB 渗漏较 NS 组明显增加;低剂量 CHR 预处理组(C)及高剂量 CHR 预处理组(D)随 CHR 剂量增加,脑实质中红色荧光逐渐减少,EB 渗漏较 LPS 组明显改善 中倍放大

**2.4 脑组织病理学变化:**HE 染色可见,NS 组脑组织结构清晰紧密(图 2A);LPS 组脑组织结构疏松,部分神经元核固缩,小血管周围间隙明显增宽,血管周围水肿明显(图 2B); $C_L$ +LPS 组及  $C_H$ +LPS 组血管周围间隙较 LPS 组减小,水肿改善,以  $C_H$ +LPS 组改善更明显(图 2C ~ 2D)。

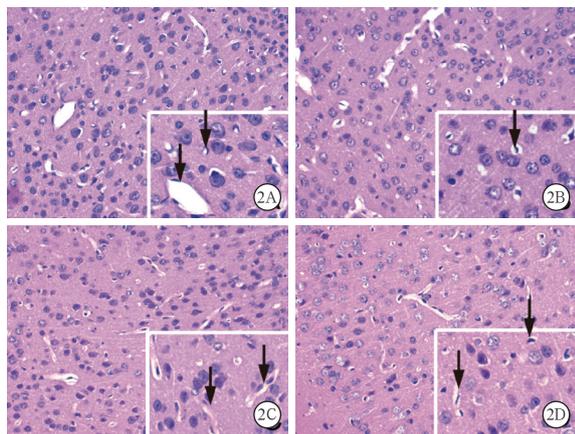


图2 光镜下观察嗜铬粒蛋白A衍生多肽CGA<sub>47-66</sub>(CHR)对脂多糖(LPS)诱导脓毒症小鼠脑皮质病理学变化的影响。箭头所示为血管周围间隙。生理盐水对照组(NS组,A)脑血管结构清晰致密,周围间隙未见明显增大;LPS组(B)脑组织结构疏松,小血管周围间隙明显增宽,血管周围水肿明显;低剂量CHR预处理组(C)及高剂量CHR预处理组(D)可见血管结构疏松,血管周围水肿明显,但较LPS组明显改善。HE染色,中倍放大,框内为高倍放大。

### 3 讨论

血脑屏障是维持脑组织细胞外环境的重要自然屏障,可选择性地允许血液中的物质进入脑组织,脑微血管内皮细胞是其最重要的组成成分。脓毒症炎症状态可导致脑微血管内皮细胞损伤<sup>[13]</sup>,引起血脑屏障通透性增加,促炎性介质及神经毒性物质通过血脑屏障进入脑实质,引起脑内环境紊乱和脑水肿。

本研究中经腹腔注射10 mg/kg LPS诱导的脓毒症小鼠在3 h、6 h后脑含水量明显增加,随后逐渐下降;6~24 h脑组织EB含量呈时间依赖性升高,提示脓毒症早期即出现血脑屏障破坏,血管内水分及大分子物质可透过血管壁渗出到脑组织。Liu等<sup>[14]</sup>通过静脉注射30 mg/kg LPS诱导脓毒症大鼠1 h后,脑MRI信号强度出现异常,血脑屏障紧密连接超微结构改变,提示血脑屏障破坏和脑水肿。Rosengarten等<sup>[15]</sup>采用静脉注射5 mg/kg LPS建立脓毒症大鼠模型3.5 h后,脑MRI并未显示有水肿表现。由此我们推测,LPS诱导脓毒症动物模型脑水肿出现的时间及程度与动物种类、脓毒症制模手段及LPS剂量有关。本实验中,小鼠脑EB含量增加晚于脑水肿,表明脓毒症时血液中大分子物质与水分的渗出和吸收并不完全同步。本研究中制模后6 h脑含水量和EB含量均发生变化,故选取6 h作为观察CHR对血脑屏障影响的时间点。

CGA是神经内分泌细胞嗜铬性颗粒内合成分

泌的重要激素原蛋白,与儿茶酚胺共储存、共分泌。本课题组前期研究发现,脓症患者血清CGA水平远远超过健康者,且CGA水平与疾病严重程度及预后密切相关<sup>[16]</sup>。CGA衍生多肽具有多种重要的生理功能。研究表明,CGA<sub>1-76</sub>具有多种心血管活性<sup>[17]</sup>,而CGA<sub>47-66</sub>(即CHR)是CGA<sub>1-76</sub>的核心基团,具有抗真菌活性,能穿透细胞膜。本课题组在体外实验中证实,CHR能改善炎性介质引起的血管内皮细胞通透性增加<sup>[12]</sup>。本研究中观察到,CHR在活体内进行预处理能减少LPS致脓毒症模型小鼠脑组织血管内水分和大分子物质渗漏,对血脑屏障功能具有明显的改善作用,并呈一定量效关系,与体外实验得出“CHR可呈剂量依赖性保护血管内皮细胞”的结果一致<sup>[18]</sup>。

脓毒症时机体通过全身炎症反应激活下丘脑-垂体-肾上腺皮质轴、蓝斑/交感神经-肾上腺髓质轴及胆碱能抗炎通路<sup>[19-22]</sup>,调动全身多个系统共同参与发生发展过程。研究表明,在这个应激过程中,上调的糖皮质激素和胆碱能激素能发挥抗炎作用<sup>[23-25]</sup>。CGA作为应激激素的功能尚未阐明<sup>[26]</sup>,本研究证实CGA衍生多肽CHR对脓毒症血脑屏障通透性紊乱具有调节作用,从神经内分泌调节的角度阐述脓毒症时血脑屏障稳态调节和器官功能的自身保护机制,为SAE的防治提供了新思路。

本研究也存在不足:通过腹腔注射CHR而未采用静脉注射的方式,因此小鼠腹膜对CHR的吸收效率可能影响实验结果;未对CHR有效血药浓度进行检测,也未探索其最佳应用浓度。

综上所述,脓毒症早期即可出现血脑屏障通透性改变和脑水肿,而CGA衍生多肽CHR可以改善脓毒症时血脑屏障的高通透性,改善脑水肿,从而对SAE起到保护作用。本研究首次证实了CGA通过维持血脑屏障稳态在脓毒症中起到抗炎作用。

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## • 读者 • 作者 • 编者 •

## 本刊对基金目标注的有关要求

基金项目指论文产出的资助背景, 例如国家自然科学基金、国家高技术研究发展计划 (863 计划)、国家科技攻关计划、国家重点基础研究发展计划 (973 计划); 行业专项基金列出提供基金的单位, 如国家卫生和计划生育委员会科研基金, 临床重点专项基金建设项目可只列出国家临床重点学科建设项目、中医药管理局临床重点学科建设项目或各省市自治区临床重点专科建设项目等。各省市基金标注方法同上。基金项目采用双语著录, 分别置于中、英文摘要关键词下方。

示例:

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