

## • 论著 •

# 膜攻击复合物 C5b-9 对创伤失血性休克大鼠肝损害的影响研究

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**【摘要】目的** 观察大鼠创伤失血性休克时肝脏是否受膜攻击复合物攻击,以及膜攻击复合物是否对肝细胞凋亡产生影响。**方法** 雄性 Wistar 大鼠 50 只,按随机数字表法均分为正常组及模型 1、3、6、24 h 组 5 组。采用骨折后经颈动脉放血至血压 40 mm Hg(1 mm Hg=0.133 kPa)制备创伤失血性休克模型。取血浆,采用酶联免疫吸附法(ELISA)检测膜攻击复合物 C5b-9 浓度,采用速率法测定丙氨酸转氨酶(ALT)、天冬氨酸转氨酶(AST)浓度;取肝脏组织,采用免疫组化法检测 C5b-9 阳性表达,用原位末端缺刻标记法(TUNEL)检测肝细胞凋亡,苏木素-伊红(HE)染色,光镜下观察病理改变。**结果** 正常组血中能检测到少量的 C5b-9;模型 1、3、6 h 组血中 C5b-9 浓度(ng/L)较正常组显著升高( $272.91 \pm 9.56$ 、 $192.01 \pm 9.04$ 、 $156.78 \pm 8.37$  比  $25.98 \pm 5.87$ , 均  $P < 0.05$ )。模型 3 h 组血 ALT(U/L)、模型 1 h 组血 AST(U/L)即显著上升( $92.90 \pm 8.83$ 、 $264.83 \pm 31.14$ ),24 h 达高峰( $184.30 \pm 12.98$ 、 $647.36 \pm 60.02$ ),与正常组( $38.75 \pm 5.40$ 、 $66.69 \pm 19.95$ )比较差异均有统计学意义(均  $P < 0.05$ )。正常组及模型 1 h、6 h 组肝脏未发现 C5b-9 阳性表达;模型 3 h 组门管区大量肝脏实质细胞存在 C5b-9(个)沉积( $22.60 \pm 1.06$ ),模型 24 h 组 C5b-9 沉积明显减少( $2.20 \pm 0.60$ ,  $P < 0.05$ )。正常组未检测到凋亡细胞;模型 1、6、24 h 组发现散在凋亡细胞(个: $1.20 \pm 0.25$ 、 $5.60 \pm 0.37$ 、 $1.60 \pm 0.26$ ),模型 3 h 组中央静脉周围凋亡细胞明显增加,出现凋亡高峰( $20.60 \pm 0.47$ ),与其余各组比较差异均有统计学意义(均  $P < 0.05$ )。模型组可见肝细胞水肿变性、肝细胞膜完整性破坏,细胞溶解,以 24 h 病理损害最重。**结论** 创伤失血性休克时大鼠肝脏受到膜攻击复合物 C5b-9 的攻击,并且肝脏 C5b-9 的表达高峰与凋亡高峰在同一时间点出现,但并不是同一部位;模型 3 h 后血中低水平的 C5b-9 提示预后极差。

**【关键词】** 休克, 创伤失血性; 肝脏; 膜攻击复合物; 凋亡

## The effect of the membrane attack complex C5b-9 on liver cells during traumatic hemorrhagic shock in rat

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**【Abstract】Objective** To observe whether the membrane attack complex C5b-9 would accumulate in the rats' liver after receiving the assault of traumatic hemorrhagic shock, and whether the membrane attack complex deals an impact on liver apoptosis. **Methods** Fifty male healthy Wistar rats were randomly divided into five groups: normal group, 1, 3, 6, 24-hour model groups. The model of traumatic hemorrhagic shock was reproduced by withdrawal of blood from carotid artery after a bone fracture till the blood pressure lowered to 40 mm Hg (1 mm Hg=0.133 kPa). Plasma membrane attack complex C5b-9 concentration was assayed using enzyme linked immunoabsorbent assay. Alanine aminotransferase (ALT) and aspartate aminotransferase (AST) in blood was determined by Rate method. Immunohistochemistry was used to detect C5b-9 deposition in the liver. Apoptosis of liver cells was then detected by the terminal deoxynucleotidyl transferase-mediated dUTP nick end labeling (TUNEL) assay. The pathological changes in paraffin sections stained with hematoxylin-eosin (HE) were observed under light microscope. **Results** A small amount of C5b-9 in plasma was found in normal group, and the values (ng/L) of 1, 3, 6-hour models were significantly higher than those of the normal group ( $272.91 \pm 9.56$ ,  $192.01 \pm 9.04$ ,  $156.78 \pm 8.37$  vs.  $25.98 \pm 5.87$ , all  $P < 0.05$ ). ALT (U/L) in 3-hour model group and AST (U/L) in 1-hour model group were increased significantly ( $92.90 \pm 8.83$ ,  $264.83 \pm 31.14$ ), peaked at 24 hours ( $184.30 \pm 12.98$ ,  $647.36 \pm 60.02$ ), and there was significant difference compared with normal group ( $38.75 \pm 5.40$ ,  $66.69 \pm 19.95$ , all  $P < 0.05$ ). In the normal group and the 1-hour and 6-hour model groups, no C5b-9 was found in liver, but in the 3-hour model group a large number of liver parenchymal cells in the portal area were found to contain C5b-9 ( $22.60 \pm 1.06$ ), however the number decreased significantly in the 24-hour model ( $2.20 \pm 0.60$ ,  $P < 0.05$ ). In normal group there was no apoptotic cell, and in 1, 6, 24-hour model groups there were scattered apoptotic cells ( $1.20 \pm 0.25$ ,  $5.60 \pm 0.37$ ,  $1.60 \pm 0.26$ ). In the 3-hour model group apoptosis of hepatic cells around the central vein was increased to the peak ( $20.60 \pm 0.47$ ), and there was significant difference compared with other groups (all  $P < 0.05$ ). In the model groups the liver cells became edematous, and the integrity of the membrane was lost, and some cells were even lysed. The pathological damage is most serious in 24-hour model group. **Conclusion** The membrane attack complex C5b-9 insulted the rats' liver after a

traumatic hemorrhagic shock, and apoptosis of hepatic cells and the content of C5b-9 peaked in 3-hour model, though they do not occur in the same site. A low level of C5b-9 in blood 3 hours after shock predict a poor prognosis.

**【Key words】** Traumatic hemorrhagic shock; Liver; Membrane attack complex; Apoptosis

创伤失血性休克时存在补体的大量活化<sup>[1]</sup>, 补体系统是人体重要的天然免疫防御系统之一, 由30余种血浆糖蛋白组成, 有复杂的蛋白质级联反应系统和精细的调控机制。多种病原微生物及抗原抗体复合物等可通过替代、经典和甘露聚糖结合凝集素途径(MBL)3条既交叉又独立的途径活化补体, 最终通过C5b、C6、C7、C8和C9顺序组装形成膜攻击复合物C5b-9, 造成对机体的损害。本实验中探讨了创伤失血性休克大鼠肝脏是否受膜攻击复合物的攻击以及膜攻击复合物是否与肝脏细胞凋亡有关。

## 1 材料与方法

**1.1 动物与分组:**选用二级健康雄性Wistar大鼠50只, 体重(220±30)g, 由中国军事医学科学院提供(动物合格证号:0010143)。按随机数字表法分为正常组及模型1、3、6和24 h组, 每组10只。

**1.2 创伤失血性休克模型的制备以及标本制备:**参考张匀等<sup>[2]</sup>的改良休克模型制备创伤失血性休克大鼠模型。腹腔注射水合氯醛麻醉大鼠, 用钳夹致双侧股骨、胫骨及肱骨闭合性骨折后, 以20号动静脉留置针行左侧颈动脉和右侧股静脉置管(置管前用肝素预冲)。骨折后15 min经颈动脉插管迅速放血, 使血压降至40 mm Hg (1 mm Hg=0.133 kPa), 持续60 min后回输血液并加2倍失血量的林格液复苏。于复苏后1、3、6、24 h麻醉动物取肝脏, 部分组织用中性甲醛溶液固定, 石蜡包埋、切片, 用于免疫组化和原位末端缺刻标记法(TUNEL)染色。血液经低温离心后收集血浆, -80℃保存备检。实验过程中动物处置方法符合动物伦理学标准。

## 1.3 检测指标及方法

**1.3.1 血浆C5b-9浓度检测:**采用酶联免疫吸附法(ELISA), 操作按试剂盒(购自美国RD公司)说明书进行。依次在酶标孔中滴加不同浓度标准品, 其余孔中滴加样品, 在酶标仪上波长450 nm处自动绘制标准品的标准曲线, 由酶标仪自动计算出各样品的C5b-9浓度。

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**1.3.2 血浆丙氨酸转氨酶(ALT)、天冬氨酸转氨酶(AST)浓度检测:**应用全自动生化仪, 采用速率法测量血浆ALT、AST浓度, 操作按试剂盒(购自瑞士罗氏公司)说明书进行。

**1.3.3 肝脏C5b-9表达检测:**免疫组化采用过氧化物酶标记的链霉卵白素(SP)一步法步骤进行测定。一抗C5b-9抗体购自荷兰Hyclut公司, 抗体1:60稀释, 4℃冰箱过夜孵育, 其余操作步骤按SP试剂盒(购自北京中杉金桥生物公司)说明书进行。石蜡包埋的病理切片经3,3'-二氨基联苯胺(DAB)显色, 细胞质呈棕色的细胞为阳性细胞。采用病理图像分析仪, 每组选10张切片, 每张切片随机选5个高倍非重叠视野, 记录阳性细胞数, 取均值。

**1.3.4 细胞凋亡检测:**采用TUNEL法原位标记DNA片段检测凋亡细胞, 操作按试剂盒(购自美国Promega公司)说明书进行。DAB显色, 细胞质呈棕色时为凋亡细胞。每组选10张切片, 每张切片随机选5个高倍非重叠视野, 记录凋亡细胞数, 取均值。

**1.3.5 病理观察:**石蜡包埋的病理切片脱水后常规行苏木素-伊红(HE)染色, 中性树胶封片, 光镜下观察病理改变。

**1.4 统计学处理:**应用SPSS 16.0统计软件, 数据以均数±标准差( $\bar{x} \pm s$ )表示, 组间比较采用单因素方差分析(one-way ANOVA) LSD检验,  $P < 0.05$ 为差异有统计学意义。

## 2 结果

**2.1 血浆C5b-9浓度(表1):**正常组血浆中能够检测到少量的C5b-9。制模后1 h血浆C5b-9浓度即明显增高, 随后逐渐下降, 3 h和6 h仍高于正常组(均 $P < 0.05$ ), 24 h下降至低于正常组( $P > 0.05$ );模型3、6、24 h组血浆C5b-9浓度明显低于模型1 h组(均 $P < 0.05$ )。

**2.2 血浆ALT、AST浓度(表1):**制模后3 h血浆ALT以及1 h血浆AST浓度即较正常组明显升高(均 $P < 0.05$ ), 之后逐渐升高至24 h达峰值。

**2.3 肝组织C5b-9表达(表1;彩色插页图1):**正常组及模型1 h、6 h组肝组织中未检测到C5b-9阳性表达;模型3 h、24 h组有C5b-9阳性表达, 且模型3 h组表达明显高于24 h组( $P < 0.05$ )。

**2.4 肝组织细胞凋亡(表1;彩色插页图2):**正常组

表1 创伤失血性休克大鼠不同时间点血浆C5b-9、ALT、AST浓度及肝脏C5b-9阳性表达和凋亡细胞数的变化(±s)

组别	动物数	血 C5b-9(ng/L)	血 ALT(U/L)	血 AST(U/L)	肝脏 C5b-9(个)	凋亡细胞数(个)
正常组	10	25.98±5.87	38.75±5.40	66.69±19.95	0	0
模型1 h组	10	272.91±9.56 <sup>a</sup>	59.41±8.68	264.83±31.14 <sup>a</sup>	0	1.20±0.25
模型3 h组	10	192.01±9.04 <sup>ab</sup>	92.90±8.83 <sup>ab</sup>	405.73±56.87 <sup>a</sup>	22.60±1.06	20.60±0.47 <sup>b</sup>
模型6 h组	10	156.78±8.37 <sup>ab</sup>	119.82±8.31 <sup>abc</sup>	495.11±70.78 <sup>ab</sup>	0	5.60±0.37 <sup>c</sup>
模型24 h组	10	11.78±2.73 <sup>b</sup>	184.30±12.98 <sup>abc</sup>	647.36±60.02 <sup>ab</sup>	2.20±0.60 <sup>c</sup>	1.60±0.26 <sup>c</sup>

注:C5b-9:膜攻击复合物,ALT:丙氨酸转氨酶,AST:天冬氨酸转氨酶;与正常组比较,<sup>a</sup>P<0.05;与模型1 h组比较,<sup>b</sup>P<0.05;与模型3 h组比较,<sup>c</sup>P<0.05

未发现凋亡细胞。制模后1 h组即发现散在凋亡细胞,模型3 h组凋亡细胞显著增加达峰值,随时间延长凋亡细胞数逐渐减少,模型6 h、24 h组凋亡细胞明显少于3 h组(均P<0.05)。

**2.5 肝组织病理变化(彩色插页图3):**模型组可见肝细胞水肿变性、肝细胞膜完整性破坏,细胞溶解,以24 h病理损害最重。

### 3 讨论

严重创伤性休克可引发全身炎症反应综合征(SIRS)和多器官功能障碍综合征(MODS),甚至多器官功能衰竭(MOF)。创伤时存在补体的大量活化<sup>[3]</sup>,目前关于补体在创伤失血性休克时肺脏及肠道损伤机制研究较多<sup>[4-5]</sup>。本研究中拟从补体角度研究创伤失血性休克时肝脏的损伤机制。

肝脏是产生补体的重要器官,其表面有许多补体抑制因子,因此很少受到膜攻击复合物C5b-9的攻击。目前关于C5b-9对肝脏损害机制研究主要集中在移植时的病理生理变化<sup>[6]</sup>。在肝脏缺血/再灌注(I/R)时发现C5b-9沉积于肝实质细胞,采用基因敲除方法阻断C5b-9的产生,可以改善I/R损害<sup>[7]</sup>,故认为C5b-9与移植后患者的血流动力学恶化密切相关<sup>[8]</sup>。本研究发现:模型大鼠肝脏受到C5b-9的攻击,且主要在模型3 h时门管区肝脏实质细胞;休克时肝脏细胞大量凋亡,凋亡时间与文献<sup>[9]</sup>报道的结果吻合,该文献发现正常肝脏可见散在凋亡细胞,而本实验中未发现正常肝脏细胞凋亡,可能与实验方法的敏感性差异有关;同时肝细胞凋亡主要出现在模型早期,以3 h和6 h明显,在24 h未发现明显的凋亡,且主要出现在小叶中央静脉周围的肝脏实质细胞,并存在凋亡细胞散在分布的现象。

有研究证实亚溶解的C5b-9在组织细胞的沉积可诱导凋亡的产生<sup>[10]</sup>。采用基因敲除C6方法,可发现肾脏实质细胞凋亡减少,并能显著改善肾功能<sup>[11-12]</sup>。补体通过裂解和活化天冬氨酸特异性半胱氨酸蛋白酶3(caspase-3)来诱导凋亡,利用caspase抑制剂可以完全抑制由补体诱导的凋亡<sup>[13]</sup>,而且凋

亡又可以活化补体<sup>[14]</sup>。在本研究中同时发现模型组C5b-9沉积以及肝脏实质细胞凋亡的发生,二者之间是否有必然联系,目前机制仍不是很清楚。

本实验结果显示C5b-9并没有直接造成其沉积部位细胞凋亡,而可能是通过促进诱导凋亡的其他因子分泌造成邻近细胞的凋亡。C5b-9是如何引起邻近细胞凋亡的呢?目前关于膜攻击复合物引起细胞凋亡研究如下:①C5b-9能够通过促进前炎症因子如前列腺素E、白细胞介素-1和肿瘤坏死因子(TNF)的表达,扩大炎症反应<sup>[15]</sup>。研究显示,TNF参与肝损伤的病理过程,其主要通过与肝细胞膜上的受体TNFRI结合而诱发肝细胞凋亡<sup>[16]</sup>。②可能促进凋亡诱导因子(AIF)表达,以促进线粒体外凋亡发生<sup>[17]</sup>。Fas和FasL作为凋亡的线粒体外途径-死亡受体途径,在急性肝损伤时肝脏Fas和FasL表达显著增加,与肝细胞凋亡变化相一致<sup>[18]</sup>。③C5b-9促视网膜色素上皮细胞转化生长因子-β(TGF-β)表达<sup>[19]</sup>,TGF-β可以促进凋亡<sup>[20]</sup>。④C5b-9可使人类单核细胞释放活性氧<sup>[21]</sup>,促氧化物质增多和抗氧化物质减少的氧化应激,与肝脏细胞凋亡程度呈正相关<sup>[22]</sup>。还可能与降低还原型谷胱甘肽促进了肝细胞凋亡有关<sup>[23]</sup>。⑤C5b-9也有可能通过提高活化转录因子3(ATF3)的表达促进细胞凋亡<sup>[24]</sup>。Roos等<sup>[25]</sup>对终末补体复合物诱导凋亡的机制提出了几种假说:①胞内钙离子和磷离子或神经酰胺产物的增多与凋亡信号的直接活化密切相关。同时国内学者发现,肝脏实质细胞内钙离子浓度与肝脏实质细胞的凋亡程度呈正相关<sup>[26]</sup>。②小分子复合物如一氧化氮(NO)、过氧化氢(H<sub>2</sub>O<sub>2</sub>)等可能参与诱导凋亡。国内研究证实补体能活化库普弗细胞<sup>[27]</sup>,封闭库普弗细胞后肠I/R模型中肝细胞凋亡明显增多<sup>[28]</sup>。③亚溶解的膜攻击复合物MAC的攻击与通过Thy21抗原激发所诱导的信号协同作用结果。

有研究认为,血浆中低水平的可溶性C5b-9预示心功能失调,可能与可溶性C5b-9在梗死心肌的沉积和活化有关<sup>[29]</sup>。本实验发现,模型3 h后血浆

C5b-9 越低,相应时间点 AST 越高,病理损害越重,故模型 3 h 后血中低水平 C5b-9 提示预后极差。

目前尚无关于创伤失血性休克时肝脏是否受膜攻击复合物攻击的研究报道。本研究为创伤失血性休克时肝脏损害机制研究开辟了一条新的路径,同时膜攻击复合物是否可以引起未被攻击细胞的凋亡,以及如何应用补体抑制剂等问题还需进一步研究。中药作为我国国粹,在治疗肝损伤方面研究取得了很大成效,如红花注射液和甘草提取物甘利欣可进一步研究其是否影响补体,调节免疫,从而发挥对创伤失血性休克的肝脏保护作用<sup>[30-31]</sup>。

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# 膜攻击复合物C5b-9对创伤失血性休克大鼠肝损害的影响研究

(正文见 158 页)

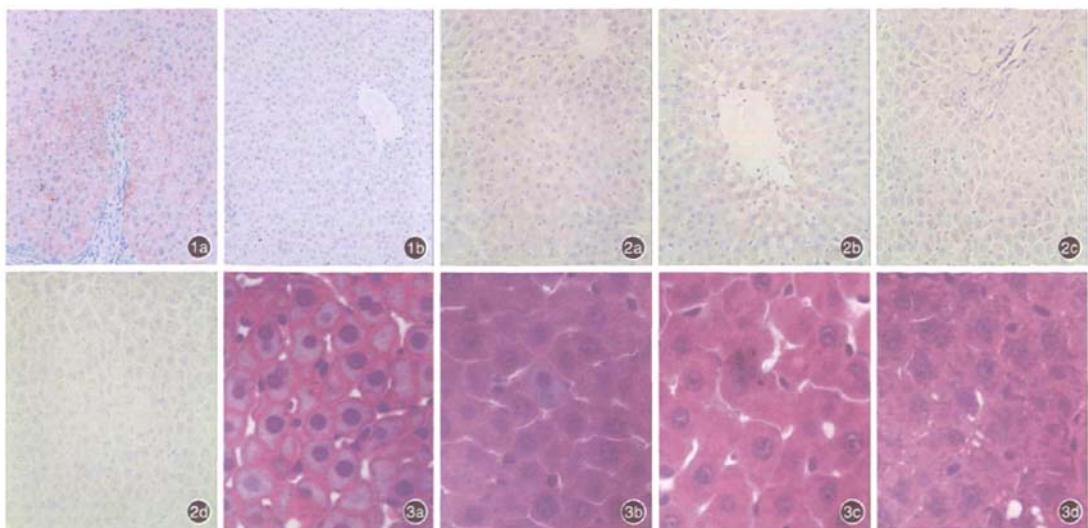


图1 光镜下观察创伤失血性休克大鼠制模后肝脏膜攻击复合物C5b-9阳性表达 细胞质中棕染部分为阳性细胞；模型3 h组(a)肝脏门管区有大量C5b-9阳性细胞沉积；模型24 h组(b)中央静脉周围的散在肝实质细胞呈淡棕染色 免疫组化 $\times 10$

图2 光镜下观察创伤失血性休克大鼠制模后肝脏细胞凋亡情况 呈棕染的细胞为凋亡细胞；模型1 h组(a)有少量凋亡细胞，个别细胞呈淡棕染色；模型3 h组(b)凋亡细胞明显增多，主要出现在小叶中央静脉周围的肝脏实质细胞，沿肝索分布；模型6 h组(c)凋亡细胞散在分布于门静脉周围的肝实质细胞；模型24 h组(d)个别细胞呈淡棕染色 TUNEL $\times 200$

图3 光镜下观察创伤失血性休克大鼠制模后肝组织病理改变 模型1 h组(a)肝细胞索清晰、结构完整，出现细胞水肿；模

型3 h组(b)肝细胞结构完整性破坏，包膜破裂，细胞间隙可见炎性细胞浸润，仍可见个别肝细胞水肿；模型6 h组(c)肝

细胞结构完整性破坏，包膜破裂，细胞融合，核膜尚完整；模型24 h组(d)肝细胞索、肝窦结构消失，肝细胞结构完整性破

坏，包膜破裂，细胞融合成片，细胞核碎裂，炎性细胞浸润增多 HE $\times 400$

# 羟乙基淀粉对脑缺血/再灌注大鼠颅内压及血浆胶体渗透压影响

(正文见 166 页)

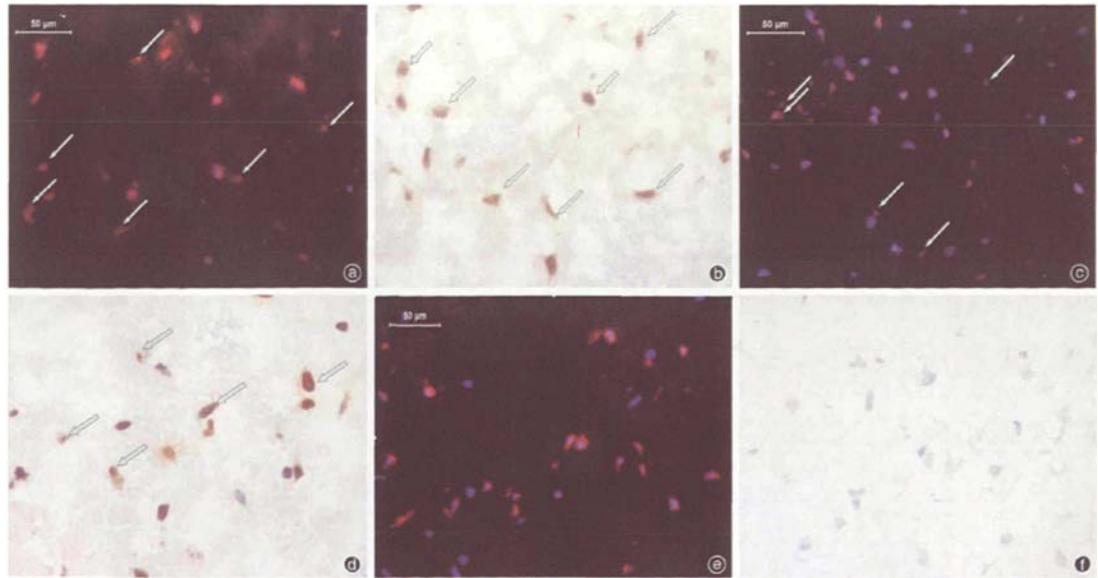


图1 光镜下观察羟乙基淀粉对脑缺血/再灌注大鼠神经元凋亡的影响 羟乙基淀粉组(a、b)可见数量不等、散在的凋亡神经元；免疫荧光4,6-二氨基-2-苯基吲哚染色(DAPI染色)可见凋亡神经元细胞核呈蓝色，红色为神经元特异核蛋白，箭头所指为凋亡后DAPI染色黯淡，崩解的神经元细胞核(a,  $\times 400$ )；原位末端缺口标记法(TUNEL)染色可见较多神经元细胞内存在凋亡小体(棕色斑块)，蓝色为神经元特异核蛋白标记的神经元细胞核(b,  $\times 400$ )。模型组(c为DAPI染色,d为TUNEL染色,  $\times 400$ )与羟乙基淀粉组(a、b)凋亡情况类似，假手术组(e为DAPI染色,f为TUNEL染色,  $\times 400$ )凋亡细胞极为罕见

## 膜攻击复合物C5b-9对创伤失血性休克大鼠肝损害的影响研究

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